

NIH Public Access

Author Manuscript

Neuropharmacology. Author manuscript; available in PMC 2013 November 01.

Published in final edited form as:

Neuropharmacology. 2012 November ; 63(6): 1172–1181. doi:10.1016/j.neuropharm.2012.06.038.

Stimulation of adenosine receptors in the nucleus accumbens reverses the expression of cocaine sensitization and cross-sensitization to dopamine D₂ receptors in rats

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Abstract

Adenosine receptors co-localize with dopamine receptors on medium spiny nucleus accumbens (NAc) neurons where they antagonize dopamine receptor activity. It remains unclear whether adenosine receptor stimulation in the NAc restores cocaine-induced enhancements in dopamine receptor sensitivity. The goal of these studies was to determine whether stimulating A_1 or A_{2A} receptors in the NAc reduces the expression of cocaine sensitization. Rats were sensitized with 7 daily treatments of cocaine (15 mg/kg, i.p.). Following one-week withdrawal, the effects of intra-NAc microinjections of the adenosine kinase inhibitor (ABT-702), the adenosine deaminase inhibitor (deoxycoformycin; DCF), the specific A_1 receptor agonist (CPA) and the specific A_{2A} receptor agonist (CGS 21680) were tested on the behavioral expression of cocaine sensitization. The results indicate that intra-NAc pretreatment of ABT-702 and DCF dose-dependently blocked the expression of cocaine sensitization while having no effects on acute cocaine sensitivity, suggesting that upregulation of endogenous adenosine in the accumbens is sufficient to nonselectively stimulate adenosine receptors and reverse the expression of cocaine sensitization. Intra-NAc treatment of CPA significantly inhibited the expression of cocaine sensitization, which was reversed by both A1 and A2A receptor antagonism. Intra-NAc treatment of CGS 21680 also significantly inhibited the expression of cocaine sensitization, which was selectively reversed by A_{2A}, but not A₁, receptor antagonism. Finally, CGS 21680 also inhibited the expression of quinpirole cross-sensitization. Together, these findings suggest that adenosine receptor stimulation in the NAc is sufficient to reverse the behavioral expression of cocaine sensitization and that A_{2A} receptors blunt cocaine-induced sensitization of post-synaptic D₂ receptors.

Keywords

 A_1 receptor A_{2A} receptor; D_2 receptor; psychostimulant; locomotor; dopamine receptor; adenosine kinase; adenosine deaminase; pentostatin

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1. Introduction

Cocaine is a psychostimulant that can lead to dependence following continuous use, resulting in an addiction that negatively affects the individual's physical, mental, and social health. Animals display abnormal behaviors such as locomotor hyperactivity upon acute exposure to cocaine that escalate with repeated cocaine exposure, resulting in locomotor sensitization (Robinson and Berridge, 2008, Thomas et al., 2008). The enhanced locomotor response observed in animals following withdrawal and re-exposure to the same dose of cocaine is thought to correspond to behaviors such as compulsive drug seeking and relapse seen in human cocaine addicts (Kalivas et al., 1998, Robinson and Berridge, 2008). Thus, locomotor sensitization is conceptualized as an expression of cocaine-induced neurobiological alterations that may persist in periods of withdrawal and contribute to the enhanced susceptibility to relapse. A detailed understanding of the molecular and pharmacological bases of behavioral sensitization will reveal novel molecular targets for pharmacological treatment strategies to offset cocaine-induced neural alterations.

Cocaine increases synaptic levels of the neurotransmitters dopamine, serotonin, and norepinephrine by inhibiting their respective reuptake transporters (Ritz et al., 1990). A major neural circuit involved in the development and expression of cocaine sensitization is the mesolimbic DA pathway (Di Chiara, 1995). This pathway consists of dopamine cells in the ventral tegmental area that project to medium spiny GABA neurons in the nucleus accumbens (NAc). The mesolimbic dopamine pathway plays a key role in the brain's reward system and it is thought that dysregulation within this neural circuit occurs with repeated cocaine exposure (Koob, 2009, Self and Nestler, 1998). Five subtypes of receptors mediate cocaine-induced enhancements in dopamine neurotransmission, including the D₁-like receptor family consisting of D1 and D5 receptors, and the D2-like receptor family consisting of D₂, D₃, and D₄ receptors (Missale et al., 1998, Sibley et al., 1993). Rodent studies suggest that stimulation of the D₁, D₂, and D₃ receptors is critical for behavioral hyperactivity and locomotor sensitization following acute and repeated cocaine exposure (Adams et al., 2001, Kita et al., 1999, Piercey et al., 1992, Ushijima et al., 1995). Additional studies have revealed that cocaine-sensitized rodents display enhanced locomotor responses to a challenge with the selective D2-receptor agonist quinpirole, suggesting that repeated cocaine exposure sensitizes D₂ receptors (Bachtell et al., 2008, Collins et al., 2011, Edwards et al., 2007, Ujike et al., 1990). Thus, offsetting cocaine-induced D₂ receptor hypersensitivity may represent a viable strategy to reduce the expression of locomotor sensitization.

Adenosine is an endogenous purine nucleoside that acts as a neuromodulator of dopamine signaling within the mesolimbic pathway (Ferre et al., 1992, Filip et al., 2012). Extracellular adenosine can be formed in the synaptic cleft by hydrolysis of extracellular adenosine triphosphate or by passive transport from intracellular non-vesicular stores (Cass et al., 1987, Fredholm et al., 1982, Thorn and Jarvis, 1996, White, 1977). Synaptic adenosine is removed from the extracellular space by reuptake via nucleoside transporters or degradation by two key enzymes, adenosine kinase and adenosine deaminase, which convert adenosine to adenosine monophosphate and inosine, respectively (Arch and Newsholme, 1978). Pharmacological inhibition of adenosine kinase or adenosine deaminase activity leads to elevations in local extracellular adenosine concentrations, which results in enhanced nonselective stimulation of adenosine receptors (Ballarin et al., 1991, Golembiowska and Zylewska, 2000, Huber et al., 2001, Jarvis et al., 2000, Pak et al., 1994, Sciotti and Van Wylen, 1993). Adenosine receptor signaling in the striatum is primarily mediated by the A_1 and A_{2A} receptor subtypes, which are highly expressed there (Rivkees et al., 1995, Svenningsson et al., 1999b). Within the striatum, postsynaptic adenosine receptor subtypes co-localize with specific dopamine receptor subtypes in the medium spiny neurons where they appear to antagonize postsynaptic dopamine signaling (Bertran-Gonzalez et al., 2009,

Ferre et al., 1994a, Ferre et al., 1994b, Ferre et al., 1999, Gines et al., 2000, Hakansson et al., 2006, Svenningsson et al., 1998).

Emergent findings show that A_1 and A_{2A} receptors may play a critical role in regulating cocaine-induced behavioral responses (Bachtell and Self, 2009, Filip et al., 2006, Green and Schenk, 2002, Knapp et al., 2001, O'Neill et al., 2012, Poleszak and Malec, 2002b, Worley et al., 1994). Most relevant is a recent study demonstrating that systemic administration of an A2A agonist decreased, while administration of an A2A antagonist increased, cocaineinduced locomotor sensitization in rats (Filip et al., 2006). However, the specific role of striatal adenosine receptors in regulating cocaine-induced acute hyperactivity and expression of locomotor sensitization remains unclear. The current studies were designed to investigate the hypothesis that stimulation of adenosine receptors in the NAc will reverse the expression of locomotor sensitization in male Sprague Dawley rats. Specifically, we tested the effects of selective- and non-selective stimulation of adenosine receptor subtypes within the medial NAc core on the acute sensitivity to cocaine, the expression of cocaine-induced locomotor sensitization, and the expression of D₂ receptor cross-sensitization in cocaine-sensitized rats. We chose to modulate adenosine receptors in the medial NAc core given structural, neurochemical and molecular changes that occur in this subregion during behavioral sensitization and the relevance of this site for other behaviors such as reinstatement to drug seeking (Brenhouse et al., 2007, Ito et al., 2004, Li et al., 2004, McFarland and Kalivas, 2001, Pierce et al., 1996). To stimulate adenosine receptors specifically in the medial NAc core, we surgically implanted rats with guide cannulae directed at the NAc core. We administered microinjections of adenosine kinase and adenosine deaminase inhibitors to assess the effects of raising endogenous extracellular adenosine and selective A1 and A2A agonists to assess the effects of selectively stimulating these receptor subtypes, on cocaineinduced locomotion. We also used subtype-selective adenosine receptor antagonists alone and in combination with the agonists to test the receptor specificity of our effects.

2. Materials and Methods

2.1 Animals and housing conditions

Male Sprague-Dawley rats (Charles River, Wilmington, MA) initially weighing 275–325 grams were individually housed with food and water available ad libitum. All experiments were conducted during the light period of a 12-h light/dark cycle in accordance with the guidelines established by the Institutional Animal Care and Use Committee at the University of Colorado at Boulder. All efforts were made to minimize animal suffering, to reduce the number of animals used, and to utilize alternatives to in vivo techniques.

2.2 Surgical procedures

Surgical implantation of intracranial cannulae was performed under halothane anesthesia (1–2.5%). Each rat was placed into a stereotaxic instrument, the scalp incised and retracted, and the head was positioned with Bregma and Lambda at the same depth coordinate. Screws were secured into the skull and holes were drilled in order to bilaterally insert guide cannulae into the NAc core (A/P: +1.7, M/L: +/-1.5, D/V: -5.7 from bregma; (Paxinos and Watson, 1998). Once inserted, guide cannulae were fixed in place with dental cement. Dummy stylets extending 1 mm beyond the tip of the cannulae were placed into the guide cannulae to maintain patency. Rats were allowed 6–7 days recovery in their home cage before experimental procedures began.

2.3 Microinjection procedure and histology

Microinjections were administered as pretreatments 5 min prior to systemic challenge injections. All microinjections occurred in the NAc at a volume of 0.5 μ L. Infusions

occurred over a 1 min period, and the microinjectors were removed 1 min after administration of the full volume of the infusion to ensure that the surrounding tissues took up the full volume. After all experimental procedures were complete, rats were euthanized with carbon dioxide gas and 1.0 μ L/side of 0.1% cresyl violet was infused intracranially to verify cannulae tip placements. Placements were determined from coronal slices and recorded on histological maps. Data from rats with incorrect placements were excluded from these studies.

2.4 General locomotor sensitization procedure

Locomotor activity was recorded in plexiglass chambers (San Diego Instruments, San Diego, CA, USA) measuring 16x16x15 inches with 16 pairs of photobeams spaced 1 inch apart on both the×and y axes. All locomotor tests were performed in darkened chambers during the light phase of the light:dark cycle. Animals were initially habituated to the locomotor testing chambers for 2-h the day prior to start of the sensitization procedure. Saline or cocaine (15 mg/kg, i.p.) injections were performed daily for one week and horizontal locomotor activity was assessed in the activity monitoring chambers. Only animals meeting criterion for locomotor sensitization (Day 7 cocaine-induced activity > 1.4 times Day 1 cocaine-induced locomotor activity) were subsequently challenged and tested.

2.5 Effects of adenosine kinase and adenosine deaminase inhibition on expression of cocaine sensitization

After 7 daily cocaine treatments (15 mg/kg, i.p.) and home cage withdrawal (7 days), animals were habituated and given an intra-NAc pretreatment of vehicle (phosphate-buffered saline (PBS, pH 7.2), the adenosine kinase inhibitor (ABT-702; 2.5 ng/side or 5 ng/side), or the adenosine deaminase inhibitor (DCF; 10 μ g/side or 20 μ g/side) five minutes prior to the cocaine challenge treatment (15 mg/kg cocaine, i.p.). Locomotor activity was assessed for 2 h after the treatments. In order to assess the effects of ABT-702 and DCF on acute sensitivity, rats run in parallel were administered 7 daily saline treatments, followed by "withdrawal" (7 days) and tested with an intra-NAc pretreatment and acute cocaine challenge according to the above procedures. Forty-eight hours later, the effects of the ABT-702 and DCF alone were tested in a similar procedure without the systemic challenge treatment.

2.6 Effects of adenosine A_1 and A_{2A} receptor stimulation on expression of cocaine sensitization

After 7 daily treatments (saline or 15 mg/kg cocaine, i.p.) and home cage withdrawal (7 days), animals were habituated and given an intra-NAc pretreatment of vehicle (PBS, pH 7.2), the A₁ receptor agonist (CPA: 0.75 μ g/side or 1.50 μ g/side), or the A_{2A} receptor agonist (CGS 21680: 2.5 ng/side or 5 ng/side) five minutes prior to challenge treatment (15 mg/kg cocaine, i.p.). Locomotor activity was assessed for 2 h after the treatments. In order to assess the effects of CPA and CGS 21680 on acute sensitivity, rats run in parallel were administered 7 daily saline treatments, followed by "withdrawal" (7 days) and tested with an intra-NAc pretreatment and acute cocaine challenge according to the above procedures.

2.7 Effects adenosine A₁ and A_{2A} receptor blockade in cocaine-naïve and cocainesensitized animals

After 7 daily treatments (saline or 15 mg/kg cocaine, i.p.) and home cage withdrawal (7 days), animals were habituated and given an intra-NAc pretreatment of vehicle (50% DMSO), the A₁ receptor antagonist (DPCPX): 50 μ g/side), or the A_{2A} receptor antagonist (MSX-3: 2.5 ng/side or 5 ng/side). MSX-3 is a prodrug of the selective A_{2A} receptor antagonist MSX-2 that is rapidly converted to its active form by phosphatases in vivo

(Muller et al., 1998, Sauer et al., 2000), and has been shown to be suitable for intracranial microinfusion (Hauber et al., 1998). Locomotor activity was assessed for 2 h after the treatments.

2.8 Effects of adenosine A_1 and A_{2A} receptor antagonism alone and in combination with adenosine receptor agonists on the expression of cocaine sensitization

After 7 daily treatments (15 mg/kg cocaine, i.p.) and home cage withdrawal (7 days), animals were habituated and given an intra-NAc pretreatment of vehicle (DMSO), the A₁ receptor antagonist (DPCPX): 50 μ g/side), or the A_{2A} receptor antagonist (MSX-3: 2.5 ng/ side or 5 ng/side) five minutes prior to a second intra-NAc treatment of vehicle (PBS, pH 7.2), the A₁ receptor agonist (CPA: 1.50 μ g/side), or the A_{2A} receptor agonist (CGS 21680: 5 ng/side). Finally, a challenge treatment (15 mg/kg cocaine, i.p.) was administered and locomotor activity was assessed for 2 h after the treatments. Thus, all animals were sensitized to cocaine and challenged with cocaine on the test day following one of the 9 possible antagonist/agonist combinations.

2.9 Effects of adenosine A_{2A} receptor stimulation on expression of quinpirole crosssensitization

Forty-eight hours after testing the effects of adenosine A_{2A} receptor stimulation on the expression of cocaine sensitization, animals were tested for D_2 receptor cross-sensitization utilizing a 4-h within-session dose–response protocol as follows: 1-h habituation followed by hourly ascending doses of D_2 receptor agonist (saline, 0.1 and 0.3 mg/kg quinpirole, s.c.). Immediately preceding the last hour of testing and treatment with 0.3 mg/kg quinpirole, animals were administered an intra-NAc treatment with either vehicle (PBS) or CGS 21680 (5 ng/side).

2.9 Drugs

The adenosine kinase inhibitor, ABT-702 (5-(3-Bromophenyl)-7-[6-(4-morpholinyl)-3pyrido[2,3-*d*]byrimidin-4-amine dihydrochloride), the adenosine deaminase inhibitor, deoxycoformycin, ((8*R*)-3-(2-Deoxy- β -D-*erythro*-pentofuranosyl)-3,4,7,8tetrahydroimidazo[4,5-*d*][1,3]diazepin-8-ol), the A₁ receptor agonist, CPA (*N6*cyclopentyladenosine) and the A_{2A} receptor agonist, CGS 21680 [4-[2-[[6-Amino-9-(Nethyl-b-D-ribofuranuronamidosyl)-9H-purin-2-yl]amino] ethyl]benzenepropanoic acid hydrochloride] were purchased from Tocris Bioscience (Ellisville, MO). The adenosine A₁ antagonist, DPCPX (8-cyclopentyl-1,3-dipropylxanthine), the adenosine A_{2A} receptor antagonist, MSX-3 [3,7-dihydro-8-[(1E)-2-(3-methoxyphenyl)ethenyl]-7-methyl-3-[3-(phosphonooxy)propyl-1-(2-propynyl)-1H-purine-2,6-dione disodium salt hydrate], D₂selective agonist, quinpirole [(-)-Quinpirole hydrochloride], and cocaine hydrochloride were obtained from Sigma-Aldrich (St. Louis, MO). All drugs were dissolved in sterile-filtered phosphate-buffered saline (PBS, pH 7.2), except cocaine, which was dissolved in sterilefiltered physiological saline (0.9%) and DPDPX, which was dissolved in 50% DMSO in PBS.

2.10 Statistical analyses

Locomotor data (total beam breaks) were analyzed by one-way or 2-factor ANOVA with Cocaine treatment (saline or cocaine) and challenge pre-treatment (intra-NAc pretreatment dose) as the factors. Interactive effects were followed by simple main effects analyses (one-way ANOVA or t-tests) and post hoc tests (Bonferroni's test). Statistical significance was preset at p < 0.05.

3. Results

3.1 Inhibition of adenosine kinase or adenosine deaminase inhibits the expression of cocaine sensitization

All animals were administered 7 daily saline or cocaine (15 mg/kg, ip) injections to induce locomotor sensitization (Table 1). Following 7 days of withdrawal, animals were tested for the expression of cocaine sensitization in the presence or absence of the adenosine kinase inhibitor, ABT-702, or the adenosine deaminase inhibitor, DCF. Figure 1 illustrates that an intra-NAc pretreatment of either ABT-702 or DCF reduced the expression of cocaine sensitization. These effects were observed only in cocaine-sensitized animals. A significant cocaine X ABT-702 Dose interaction ($F_{2.46} = 7.47$; p = 0.0015) and significant main effects of cocaine ($F_{1.46} = 7.131$; p = 0.0096) and ABT-702 Dose ($F_{2.46} = 4.18$; p = 0.0215) were observed. Subsequent analysis of the interaction found that ABT-702 treatment in cocainenaïve animals did not significantly alter acute cocaine sensitivity ($F_{2,22} = 1.87$; p = 0.1781). However, ABT-702 treatment in cocaine-sensitized animals significantly inhibited the expression of cocaine sensitization at both ABT-702 doses ($F_{2,22} = 9.19$; p < 0.0011). Significant main effects of cocaine ($F_{1,43} = 14.73$; p = 0.0004) and DCF Dose ($F_{2,43} = 14.73$; p = 0.0004) and $F_{2,43} = 14.73$; p = 0.0004; 4.677; p = 0.0145) were also observed, however a significant cocaine X DCF Dose interaction ($F_{2,43} = 2.06$; p = 0.1394) was not. Subsequent analysis of the significant main effects found that only the high DCF dose (10 µg/side) significantly inhibited cocaine sensitivity ($F_{2.46} = 3.53$; p = 0.0376).

Figure 2 displays the effects of the adenosine kinase and adenosine deaminase inhibitors in the absence of a cocaine challenge in both cocaine-naïve and cocaine-sensitized animals. Intra-NAc administration of the inhibitors had no significant effect on locomotor activity in either cocaine-naïve or cocaine-sensitized animals.

3.2 Intra-NAc stimulation of adenosine A₁ and A_{2A} receptors blocks expression of cocaine sensitization

Inhibiting either adenosine kinase or adenosine deaminase is associated with accumulation of extracellular adenosine leading to non-selective stimulation of adenosine receptors (Ballarin et al., 1991, Golembiowska and Zylewska, 2000, Huber et al., 2001, Jarvis et al., 2000, Pak et al., 1994, Sciotti and Van Wylen, 1993). We next sought to determine whether selective stimulation of either the adenosine receptor A_1 or A_{2A} subtype in the NAc would recapitulate the inhibition of cocaine sensitization produced by enzyme inhibition. Similar to those studies, all animals were administered 7 daily saline or cocaine (15 mg/kg, ip) injections to induce locomotor sensitization (Table 2). Following 7 days of withdrawal, animals were tested for the expression of cocaine sensitization in the presence or absence of the adenosine A1 agonist, CPA, or the adenosine A2A agonist, CGS 21680. Figure 3 illustrates that an intra-NAc pretreatment of either CPA or CGS 21680 reduced the expression of cocaine sensitization. Like the enzyme inhibitors, these effects were observed only in cocaine-sensitized animals. A significant cocaine X CPA Dose interaction ($F_{2,27}$ = 3.65; p = 0.0395) and significant main effects of cocaine ($F_{1,27} = 42.27$; p = 0.0001) and CPA Dose ($F_{2,27} = 4.533$; p = 0.0201) were observed. Subsequent analysis of the interaction found CPA treatment in cocaine-naïve animals did not significantly alter acute cocaine sensitivity ($F_{2,9} < 1$; p = 0.8771) while CPA treatment in cocaine-sensitized animals significantly inhibited the expression of cocaine sensitization at both CPA doses ($F_{2.18}$ = 7.84; p = 0.0036). Similarly, a significant cocaine X CGS 21680 Dose interaction ($F_{2,25}$ = 11.23; p = 0.0003) and significant main effects of cocaine ($F_{1.25} = 52.53$; p < 0.0001) and CGS 21680 Dose ($F_{2,25} = 17.84$; p < 0.0001) were observed. Subsequent analysis of the interaction shows that CGS 21680 treatment in cocaine-sensitized animals significantly inhibited the expression of cocaine sensitization at both CGS21680 doses ($F_{2,17} = 33.06$; p <

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0.0001) and had no effect on cocaine sensitivity in cocaine-naïve animals ($F_{2,14} < 1$; p = 0.4498).

3.3 Intra-NAc blockade of adenosine A₁ and A_{2A} receptors reverses agonist-induced reductions of expression of cocaine sensitization

Previous work demonstrates that systemic administration of an adenosine A_{2A} antagonist enhances the expression of cocaine sensitization (Filip et al., 2006). Therefore, we sought to determine whether adenosine receptor blockade directly in the NAc core would produce a similar effect. We first tested whether intra-NAc adenosine A_1 and A_{2A} receptor blockade would produce cross-sensitization in cocaine-sensitized animals compared to cocaine-naïve animals. Administration of the A_1 and A_{2A} receptor antagonists produced modest increases in locomotor activity in cocaine-sensitized animals compared to vehicle treatments (Figure 4). A significant main effect of cocaine ($F_{1,26} = 5.277$; p = 0.0259) and antagonist ($F_{2,26} =$ 5.581; p = 0.0119) was observed, however the cocaine X antagonist interaction was not statistically significant ($F_{2,26} = 1.256$; p = 0.3015).

Given that adenosine receptor blockade produced only modest effects when administered alone, we next tested the specificity of the effects observed with intra-NAc administration of adenosine agonists. Similar to those studies, animals were administered 7 daily cocaine (15 mg/kg, ip) injections to induce locomotor sensitization. Following 7 days of withdrawal, animals were tested for the expression of cocaine sensitization with a cocaine challenge (15 mg/kg, ip). Prior to the cocaine challenge, animals received intra-NAc treatments of the adenosine A_1 or A_{2A} antagonist alone or in combination with intra-NAc treatments of the A_1 or A_{2A} agonists. Figure 5 illustrates the results of the antagonist/agonist combinations on the expression of cocaine sensitization. A significant agonist (vehicle, CPA, CGS 21680) X antagonist (vehicle, DPCPX, MSX-3) interaction ($F_{4,37} = 4.55$; p = 0.0044) and significant main effects of agonist ($F_{2,37} = 23.81$; p = 0.0001) and antagonists ($F_{2,37} = 3.78$; p = 0.0320) were observed. Subsequent analysis of the interaction found that CPA and CGS 21680 treatment alone significantly inhibited the expression of cocaine sensitization as observed above ($F_{2,12} = 22.79$; p = 0.0002). There was no effect of either antagonist pretreatment (DPCPX or MSX-3) on the expression of cocaine sensitization compared to vehicle pretreatment ($F_{2,13} < 1$; p = 0.6571). Pretreatment with DPCPX significantly reversed CPAinduced inhibition of cocaine sensitization while having no effect on CGS 21680-induced inhibition ($F_{2,14} = 18.92$; p = 0.0001). Pretreatment with MSX-3 significantly reversed CGS 21680-induced inhibition of cocaine sensitization ($F_{2,11} = 21.95$; p = 0.0001). There was a statistical trend for MSX-3 to reverse CPA-induced inhibition of cocaine sensitization, although this did not reach statistical significance ($F_{2,13} = 3.48$; p = 0.0616).

3.4 Intra-NAc stimulation of adenosine A_{2A} receptors inhibits quinpirole sensitivity

Cocaine sensitization is associated with a cross-sensitization in postsynaptic D₂ receptors in the NAc (Bachtell et al., 2008, Collins et al., 2011, Edwards et al., 2007, Ujike et al., 1990). This study tests the hypothesis that cocaine-induced cross-sensitization in D₂ receptor sensitivity will be inhibited though the stimulation of adenosine A_{2A} receptors that are colocalized with postsynaptic D₂ receptors in the NAc spiny neurons. Figure 4 illustrates that intra-NAc administration of the A_{2A} receptor agonist, CGS 21680, suppresses quinpirole-induced locomotor activity in both cocaine-naïve and cocaine-sensitized animals. Statistical analysis reveals that cocaine-sensitized animals have a more robust locomotor response to quinpirole administration (F_{1,16} = 12.21; p = 0.0030) and that an intra-NAc pretreatment of CGS 21680 suppresses quinpirole-induced activity in both cocaine-naïve and cocaine sensitized animals (F_{1,16} = 8.65; p = 0.0096). No interactive effects were observed (F_{2,16} = 1.04; p = 0.3222).

4. Discussion

These studies elucidate adenosine signaling in the NAc as a key pharmacological mechanism for counteracting the expression of cocaine-induced locomotor sensitization. It is important to note that these effects do not appear to be suppressing general locomotor activity since they have no effect on the acute sensitivity of cocaine. These findings corroborate previous studies demonstrating that systemic injections of adenosine agonists counteract many cocaine-induced behavioral changes. For example, systemic administration of adenosine agonists attenuates both the development and expression of behavioral sensitization to cocaine (Filip et al., 2006, Poleszak and Malec, 2002b), impairs the acquisition of cocaine self-administration (Knapp et al., 2001), reduces the expression of cocaine conditioned place preference (Poleszak and Malec, 2002a), and attenuates cocaine seeking (Bachtell and Self, 2009). Adenosine receptors are highly expressed in the striatal structures, including the NAc, however, there is expression elsewhere in the brain. Therefore, it is important to characterize the localization of these behavioral effects. We recently demonstrated that cocaine seeking is reduced by adenosine receptor stimulation and enhanced by adenosine receptor blockade specifically in the NAc core (O'Neill et al., 2012). Together, these findings indicate that pharmacological stimulation of adenosine receptors specifically in the NAc opposes the behavioral responsiveness to cocaine.

We utilized two approaches to study the influence of adenosine signaling on cocaine sensitization. First, we infused inhibitors of the enzymes adenosine kinase and adenosine deaminase directly into the NAc prior to a cocaine challenge. These are two primary enzymes responsible for adenosine metabolism in the CNS, and they directly regulate the extracellular concentration of adenosine (Golembiowska and Zylewska, 2000, Jarvis et al., 2000). Inhibition of these enzymes produces elevations in endogenous adenosine and subsequent non-selective stimulation of adenosine receptors (Ballarin et al., 1991, Golembiowska and Zylewska, 2000, Huber et al., 2001, Jarvis et al., 2000, Pak et al., 1994, Sciotti and Van Wylen, 1993). Studies have demonstrated that adenosine kinase is the ratelimiting enzyme in ADO metabolism, while adenosine deaminase plays a less critical role in determining extracellular ADO concentration (Lloyd and Fredholm, 1995, Phillips and Newsholme, 1979, Romanowska et al., 2007). It is therefore likely that inhibiting adenosine kinase with ABT-702 leads to a more substantial increase in extracellular ADO than inhibiting adenosine deaminase with DCF. This idea may explain why ABT-702 was more efficacious and a higher dose of DCF was required to significantly block the expression of cocaine-sensitization.

As an alternative to enzyme inhibition, we also examined the effects of direct stimulation of specific ADO receptor subtypes in the NAc. ADO receptor subtypes are differentially expressed on different cellular populations within the striatum (see below). Therefore, we were exploring whether stimulation of a specific subtype localized to a unique cell population may be more or less effective in mediating cocaine-induced locomotion. Our findings suggest that both ADO A₁ and A_{2A} receptor stimulation are sufficient to reverse the expression of cocaine sensitization, however, the mechanisms of these effects may be quite different. Importantly, we show that the effects of CGS 21680 were dependent upon A_{2A} receptor stimulation since administration of the A_{2A} receptor antagonist, MSX-3, but not the A₁ receptor antagonist, DPCPX, blocked CGS 21680-induced inhibition of cocaine sensitization where both A₁ and A_{2A} receptor antagonists reversed this inhibition. This suggests that A₁ receptor activation may have cooperative actions with A_{2A} receptors in the NAc akin to those observed with dopamine D₁ and D₂ receptors in the striatum (Bachtell et al., 2005, Hopf et al., 2003, Schmidt and Pierce, 2006, White, 1987).

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The dorsal striatum and NAc are comprised primarily of medium spiny GABA neurons that include two distinct subpopulations of output neurons. These two subpopulations are differentiated by their unique expression of cellular peptides and receptor subtypes, and unique projection targets (Aubert et al., 2000, Steiner and Gerfen, 1998). For example, neurons forming the direct pathway are dynorphin/substance P-expressing and contain dopamine D₁ and adenosine A₁ receptors, while neurons of the indirect pathway are enkephalin-expressing and contain dopamine D_2 and adenosine A_{2A} receptors (Lu et al., 1998). There is accumulating evidence suggesting that these two populations of striatal neurons play differential roles in cocaine's actions (Bertran-Gonzalez et al., 2008, Lee et al., 2006, Lobo et al., 2010, Lobo and Nestler, 2011). Our findings provide additional support for the unique contribution of these populations of neurons in cocaine-mediated behaviors. First, previous work indicates that systemic CPA induces c-Fos expression in the population of striatal GABA neurons containing A_{2A} and D_2 receptors (Karcz-Kubicha et al., 2006, Karcz-Kubicha et al., 2003). Similar to our effects on the expression of cocaine sensitization, administration of an A_{2A} receptor antagonist reduces CPA-induced c-Fos expression in the striatum (Karcz-Kubicha et al., 2003). Together, these findings suggest that A_1 receptor stimulation has the capacity to activate A_{2A}/D_2 -containing cells through the stimulation of A_{2A} receptors in the striatum which may reduce the expression of cocaine sensitization. Cocaine produces a robust cross-sensitization to the D₂ receptor agonist, quinpirole (Bachtell et al., 2008, Collins et al., 2011, Edwards et al., 2007, Ujike et al., 1990). We show that local administration of the A2A agonist, CGS 21680, in the NAc reverses D2 receptor cross-sensitization which coincides with other recent findings showing that NAc A2A receptors inhibit D2-induced reinstatement to cocaine seeking (O'Neill et al., 2012). Thus, it appears that activation of the A_{2A}/D_2 -containing cells in the striatum, either directly through A_{2A} receptor stimulation or indirectly through A₁ receptor stimulation, is important in reversing cocaine-induced enhancements in dopamine D₂ receptor sensitivity that contribute to the expression of behavioral sensitization.

Generally speaking, adenosine receptor co-localization with dopamine receptors provides complementary cellular regulation where the adenosine receptors oppose cellular signaling resulting from dopamine receptor stimulation (Canals et al., 2003, Ferre, 1997, Fuxe et al., 2003, Hillion et al., 2002, Svenningsson et al., 1999a, Svenningsson et al., 1998, Svenningsson et al., 1999b). In this way, adenosine receptor stimulation can offset excessive dopamine receptor stimulation resulting from repeated psychostimulant treatments (Bailey et al., 2008, Burechailo and Martin-Iverson, 1996, Burger and Martin-Iverson, 1994, Henry and White, 1991, Ujike et al., 1990). Adenosine and dopamine receptors can alter signaling of medium spiny GABA neurons within the striatum through a variety of mechanisms. For example, these receptors form heteromeric receptor complexes through electrostatic interactions (Canals et al., 2003, Ferre et al., 1994b, Ferre et al., 1998, Franco et al., 2007, Fuxe et al., 2003, Gines et al., 2000, Hillion et al., 2002, Torvinen et al., 2002). Heteromeric formation of adenosine and dopamine receptors renders a low-affinity state of the dopamine receptor where ligand binding is inhibited and coupling of G-proteins is diminished at the dopamine receptor (Ferre et al., 1991, Ferre et al., 1998, Fuxe et al., 1998, Hillion et al., 2002, Torvinen et al., 2002, Torvinen et al., 2005). Interestingly, cocaine treatment disrupts the expression of both the A1-D1 and A2A-D2 heteromer (Marcellino et al., 2010, Toda et al., 2003), which may underlie some of the changes in behavioral responses resulting from chronic cocaine administration. Functionally, disruption of adenosine-dopamine heteromeric complexes may enable enhanced dopamine receptor activity that contributes to behavioral sensitization. Adenosine receptor stimulation may facilitate the coupling of adenosine and dopamine receptors (Vidi et al., 2008), ultimately restoring dopamine antagonism and reversing the behavioral changes following chronic cocaine administration. However, it remains unclear whether heteromeric receptor complexes or another interactive mechanism mediate our effects since receptors that are not in heteromeric complexes still play an

antagonistic and reciprocal role in modulating cellular function (Ferre, 1997, Ferre et al., 1991).

Adenosine and dopamine receptors are coupled to opposing classes of G proteins (Dunwiddie and Masino, 2001, Lachowicz and Sibley, 1997). While D_2 receptors are coupled to inhibitory GaI proteins, the complementary A_{2A} receptor is coupled to stimulatory GaS proteins. Likewise, D_1 receptors are coupled to stimulatory GaS proteins while its complementary A_1 receptor is coupled to inhibitory GaI proteins. Thus, the complementary G-protein signaling between adenosine and dopamine receptors can have profound effects on intracellular signaling cascades and neuronal excitability (Ferre et al., 1994b, Ferre et al., 1996, Ferre et al., 1999, Schiffmann et al., 2007, Svenningsson et al., 1999a, Tozzi et al., 2007). This suggests that reciprocal regulation of downstream targets of cAMP (e.g. PKA-mediated phosphorylation targets) may play a role in the expression of cocaine sensitization. While adenosine receptors obviously play a significant role in opposing dopamine neurotransmission within the striatum, the cellular mechanisms of our effects on cocaine sensitization remain obscure and it is likely that both heteromeric receptor complexes and adenosine receptor-induced intracellular signaling contribute to the modulation of these behaviors.

It is important to recognize that adenosine receptors are expressed on other cell types in the NAc, providing other possible explanations for our results. For example, expression of A_1 and A_{2A} receptor heteromeric complexes on presynaptic glutamate terminals is involved in modulating striatal glutamate release (Ciruela et al., 2006, Ferre et al., 2008, Hettinger et al., 2001, Orru et al., 2011a, Orru et al., 2011b, Rodrigues et al., 2005). Thus, stimulation of presynaptic A_{2A} receptors increases striatal glutamate release, while stimulation of A_1 receptors produces the opposite effect (Ciruela et al., 2006, Corsi et al., 2000, Corsi et al., 1999). It seems unlikely that our findings would result from A_{2A} -induced increases in glutamate release since stimulation of AMPA receptors in the NAc produces enhanced locomotion in cocaine sensitized animals and blockade of AMPA receptors prevents the expression of cocaine sensitization (Bachtell and Self, 2008, Bell and Kalivas, 1996, Pierce et al., 1996). Thus, we suspect that A_{2A} receptor stimulation is primarily influencing postsynaptic A_{2A} receptors on medium spiny neurons. Our finding that A_{2A} stimulation inhibits quinpirole-induced locomotion in both cocaine-naïve and cocaine-sensitized animals also supports this notion.

It is plausible, however, that stimulation of A_1 receptors may inhibit the ability of cocaine to enhance extracellular glutamate during the cocaine challenge that is necessary for the behavioral expression of sensitization (Bell et al., 2000, Madayag et al., 2007, Pierce et al., 1996). Adenosine A_1 receptors are also thought to be expressed on a small percentage of dopamine terminals in the striatum (Alexander and Reddington, 1989, Borycz et al., 2007, Wojcik and Neff, 1983) where A_1 receptor stimulation inhibits depolarization-induced dopamine release (Borycz et al., 2007, Ebstein and Daly, 1982, Michaelis et al., 1979). Thus, stimulation of A_1 receptors may have a multitude of effects within the NAc that contributes to reversing the expression of cocaine sensitization including inhibition of cocaine-induced dopamine and glutamate release, inhibiting the postsynaptic signaling mediated by D_1 receptors, and/or enabling A_{2A} receptors to activate the A_{2A}/D_2 -containing striatal cells.

5. Conclusions

The results of these experiments suggest an important role of adenosine and adenosine receptors within the NAc for the behavioral expression of cocaine sensitization. We demonstrate that inhibition of two enzymes responsible for degrading extracellular

adenosine, adenosine kinase and adenosine deaminase, inhibit the behavioral expression of cocaine sensitization. Presumably, these effects are achieved through the non-selective stimulation of local adenosine receptors. This notion is supported by our findings that intra-NAc stimulation of either A_1 or A_{2A} receptors inhibits the behavioral expression of cocaine sensitization. Furthermore, we demonstrated that the effects of intra-NAc A2A stimulation are likely mediated through postsynaptic expression of A2A receptors since quinpiroleinduced cross-sensitization was also inhibited, which is thought to be mediated by sensitization in postsynaptic D_2 receptors. While the antagonistic interaction between adenosine and dopamine receptors on striatal neuronal transmission is supported by these experiments, the relative contribution of heteromeric and nonheteromeric complexes is unknown. Together, our results suggest that interactions between adenosine and dopamine receptors influence striatal signaling that mediates the expression of cocaine sensitization, but not the acute sensitivity of cocaine's effects. Finally, the results of this study illuminate the potential for adenosine receptor stimulation as an effective strategy for reversing cocaine-induced sensitization that may underlie an addicts' persistent susceptibility to relapse.

Acknowledgments

This work was supported by a United States Public Health Services Grant DA 029240 (R.K.B.), the Innovative Seed Grant program at the University of Colorado, and a University of Colorado CRCW Faculty Development Award.

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Highlights

- Elevating adenosine levels in the accumbens reverses cocaine sensitization
- A1 and A2A receptor stimulation in the accumbens reverses cocaine sensitization
- Accumbens adenosine receptor stimulation does not affect acute cocaine sensitivity
- A1-induced reversal of sensitization involves both A1 and A2A receptors
- Accumbens A_{2A} receptor stimulation reverses D2 receptor cross-sensitivity

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Figure 1. Intra-NAc administration of adenosine kinase and adenosine deaminase inhibitors attenuates the expression of cocaine sensitization

(a) Animals repeatedly treated with cocaine (7 X 15 mg/kg, ip) displayed significant expression of sensitization when tested with intra-NAc vehicle and 15 mg/kg cocaine (ip) following 7 days withdrawal compared with animals administered repeated saline. Intra-NAc administration of both the adenosine kinase inhibitor (ABT-702) and adenosine deaminase inhibitor (DCF) diminished the expression of cocaine sensitization. No effect of intra-NAc ABT-702 or DCF was observed on acute cocaine sensitivity since cocaine-induced locomotor activity was equivalent in cocaine-naïve animals. (b) Time-course of locomotor activity illustrating the last 30 min of the habituation period followed by the effects of 15 mg/kg cocaine (ip) with and without the intra-NAc pretreatment ABT-702 (5 μ g/side) or DCF (10 μ g/side) in cocaine-sensitized animals. * indicates significant from cocaine-sensitized with vehicle pretreatment (p<0.001); # indicates significant from cocaine-sensitized with vehicle pretreatment (p<0.01)

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(a) An intra-NAc treatment with the adenosine kinase inhibitor, ABT-702, had no effect on locomotion in either cocaine-sensitized or cocaine-naive animals. (b) An intra-NAc treatment with the adenosine deaminase inhibitor, DCF, had no effect on locomotion in either cocaine-sensitized or cocaine-naive animals.

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Figure 3. Intra-NAc administration of adenosine receptor agonists attenuates the expression of cocaine sensitization

(a) Animals repeatedly treated with cocaine (7 × 15 mg/kg, ip) displayed significant expression of sensitization when tested with intra-NAc vehicle and 15 mg/kg cocaine (ip) following 7 days withdrawal compared with animals administered repeated saline. Intra-NAc administration of both the adenosine A_{2A} agonist (CGS 21680) and adenosine A_1 agonist (CPA) diminished the expression of cocaine sensitization. No effect of intra-NAc CGS 21680 or CPA was observed on acute cocaine sensitivity since cocaine-induced locomotor activity was equivalent in cocaine-naïve animals. (b) Time-course of locomotor activity illustrating the last 30 min of the habituation period followed by the effects of 15 mg/kg cocaine (ip) with and without the intra-NAc pretreatment CGS 21680 (5 µg/side) or CPA (1.5 µg/side) in cocaine-sensitized animals. * indicates significant from respective cocaine-naïve group (p<0.0001); # indicates significant from cocaine-sensitized with vehicle pretreatment (p<0.001)

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Figure 4. Effects of intra-NAc treatment of $\rm A_1$ and $\rm A_{2A}$ antagonists in cocaine-naïve and cocaine-sensitized animals

Animals sensitized to cocaine with 7 daily cocaine injections (15 mg/kg, ip) displayed modest increases in locomotor activity compared to cocaine-naïve (saline treated) animals. * indicates significant from respective cocaine-sensitized, vehicle group (p<0.05)

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Figure 5. Effects of intra-NAc treatment of ${\rm A}_1$ and ${\rm A}_{2{\rm A}}$ on a denosine agonist-induced inhibition of cocaine sensitization

Animals were sensitized to cocaine with 7 daily cocaine injections (15 mg/kg, ip). As reported above, CPA and CGS alone significantly reduced the expression of cocaine sensitization (left). Intra-NAc pretreatment of the A₁ antagonist, DPCPX (middle), reversed CPA-induced reductions in cocaine sensitization, but not CGS 21680-induced reductions. Intra-NAc pretreatment of the A_{2A} antagonist, MSX-3 (right), significantly reversed CGS 21680-induced and CPA-induced reductions in the expression of cocaine sensitization. * indicates significant from vehicle-vehicle group (p<0.01); # indicates significant from respective agonist-vehicle group (p<0.05)

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Figure 6. Intra-NAc treatment of an $\rm A_{2A}$ receptor agonist reduces locomotor sensitivity of the $\rm D_2$ agonist in both cocaine-naïve and cocaine-sensitized

Locomotor activity induced by a high dose of the D_2 receptor agonist, quinpirole (0.3 mg/kg, sc) is attenuated in both cocaine-naïve and cocaine-sensitized animals. Importantly, cocaine-sensitized animals displayed crosssensitization in D_2 -induced locomotion, which was attenuated to levels similar to cocaine-naïve animals. * indicates significant from vehicle treated cocaine-naïve group (p<0.0001); # indicates significant from cocaine-sensitized with vehicle pretreatment (p<0.001)



Figure 7. Localization of infusion sites in NAc core

(a) Some infusion sites were verified by an infusion of $1.0 \,\mu$ L/side of 0.1% cresyl violet through the guide cannula following euthanasia and 40 μ m sections were analyzed for accurate placements. (b)Infusion sites were also verified by staining non-infused 40 μ m brain sections with cresyl violet. There were minimal signs of gliosis or scarring following the intra-NAc infusions. (c) Infusion sites for all animals included in the study analyses. Animals having infusion sites outside of the NAc core were eliminated from statistical analyses.

Table 1

Development of locomotor sensitization with 7 daily saline or cocaine administrations was equivalent between groups prior to intra-NAc treatment on challenge day

Effects of adapasing kings and domings inhibition			
Cocaine-naïve: Repeated Saline			
<u>Day 1</u>	<u>Day 7</u>	Intra-NAc Challenge Treatment @ Day 14	
6926 ± 496.3	7504 ± 2464	Vehicle	
6894 ± 1014.1	6057 ± 1855.3	ABT 702 (2.5 µg/side)	
6494 ± 924.1	6341 ± 1155.3	ABT 702 (5.0 µg/side)	
6473 ± 660.1	6602 ± 714.8	DCF (5 µg/side)	
6622 ± 496.3	6514 ± 2162	DCF (10 µg/side)	
Cocaine-sensitized: Repeated Cocaine			
<u>Day 1</u>	<u>Day 7</u>	Intra-NAc Challenge Treatment @ Day 14	
23871 ± 5184.2	$34801 \pm 4486.0^{\ast}$	Vehicle	
23593 ± 4234.0	35122 ± 4532.6 *	ABT 702 (2.5 µg/side)	
25789 ± 3897.7	$36662 \pm 4778.2^{*}$	ABT 702 (5.0 µg/side)	
21809 ± 2453.3	32681 ± 4621.0 *	DCF (5 µg/side)	
23699 ± 5334.0	$33822 \pm 2133.6^*$	DCF (10 µg/side)	

Data represent mean (\pm SEM) beam breaks/2 hrs.

*Statistically significant difference compared to Day 1 locomotor activity, p < 0.001

Table 2

Development of locomotor sensitization with 7 daily saline or cocaine administrations was equivalent between groups prior to intra-NAc treatment on challenge day

Effects of adenosine receptor stimulation			
Cocaine-naïve: Repeated Saline			
<u>Day 1</u>	<u>Day 7</u>	Intra-NAc Challenge Treatment @ Day 14	
6921 ± 248.9	6585 ± 625.9	Vehicle	
6712 ± 281.7	6184 ± 312.8	CGS 21680 (2.5 ng/side)	
5765 ± 201.3	6500 ± 342.6	CGS 21680 (5.0 ng/side)	
6176 ± 331.3	5680 ± 641.3	CPA (0.75 µg/side)	
6528 ± 278.0	6277 ± 302.9	CPA (1.5 µg/side)	
Cocaine-sensitized: Repeated Cocaine			
<u>Day 1</u>	<u>Day 7</u>	Intra-NAc Challenge Treatment @ Day 14	
23405 ± 5154.6	$47657 \pm 3562.4^{\ast}$	Vehicle	
25105 ± 4957.7	$48622 \pm 3752.4^{\ast}$	CGS 21680 (2.5 ng/side)	
27030 ± 7532.4	50943 ± 3553.8 *	CGS 21680 (5.0 ng/side)	
28193 ± 6854.1	$51022 \pm 3357.7^{*}$	CPA (0.75 µg/side)	
27216 ± 7212.2	$48724 \pm 4251.5^{*}$	CPA (1.5 µg/side)	

Data represent mean (\pm SEM) beam breaks/2 hrs.

* Statistically significant difference compared to Day 1 locomotor activity, p < 0.001